

# Translating Tissue-based Assays into Liquid Biopsies: The Final Frontier for mRNA Assays

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Mark Kidd, PhD, Scientific and Laboratory Director

January 24, 2024

# Disclosures

Laboratory and Scientific Director  
of Wren Laboratories  
Patent holder (NETest, PPQ)

CLIA-certified and CAP-accredited liquid biopsy-focused  
molecular diagnostic laboratory in Branford, CT

# Tissue-based Assays vs Liquid Biopsies

## Conventional Biopsy

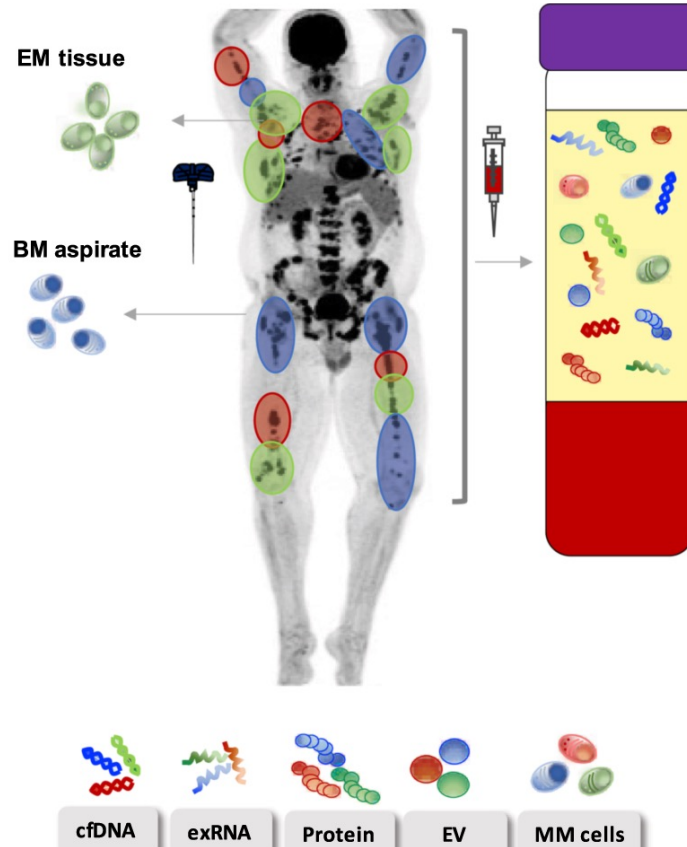
Invasive

Often unsuccessful

Limited snapshot of the tumor

No real-time evaluation

Difficult/tedious



## Liquid Biopsy

Non-Invasive

Easy to do/successful

Comprehensive tumor evaluation

Therapeutic monitoring

Scalable

**Multiple Myeloma**

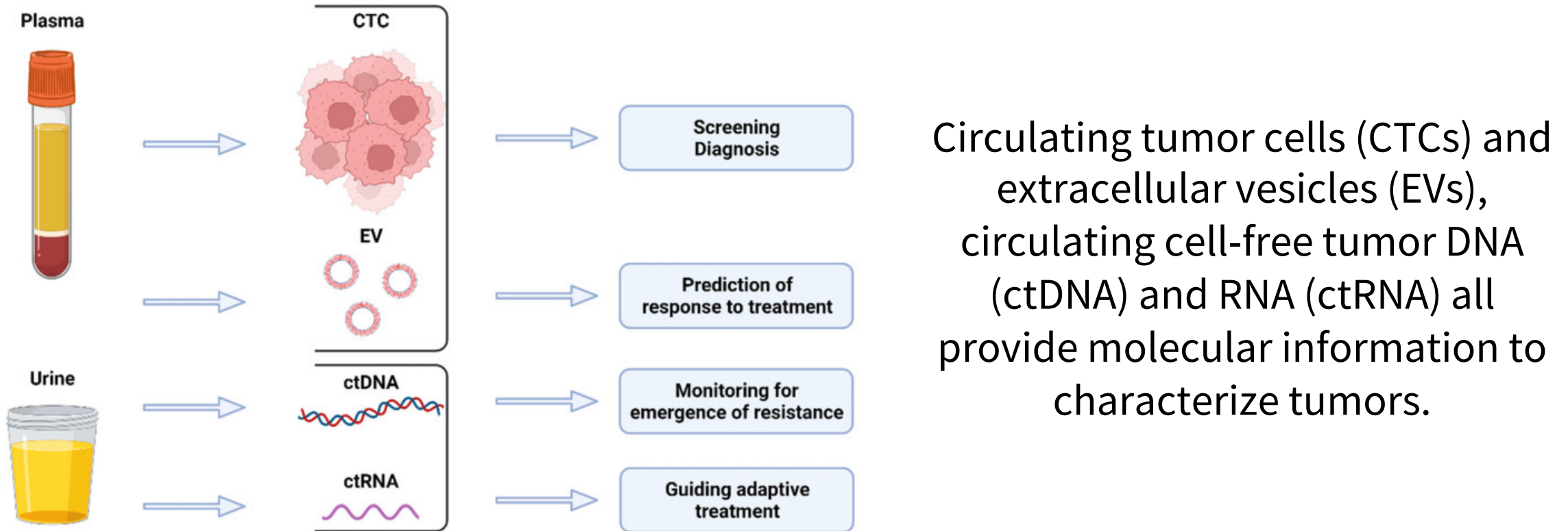
# Why Liquid Biopsy?

Traditional surgical pathology specimens (biopsies)

- Provide a snapshot of a dynamic process
- Are inadequate for monitoring
- Cannot provide necessary information to guide decision making



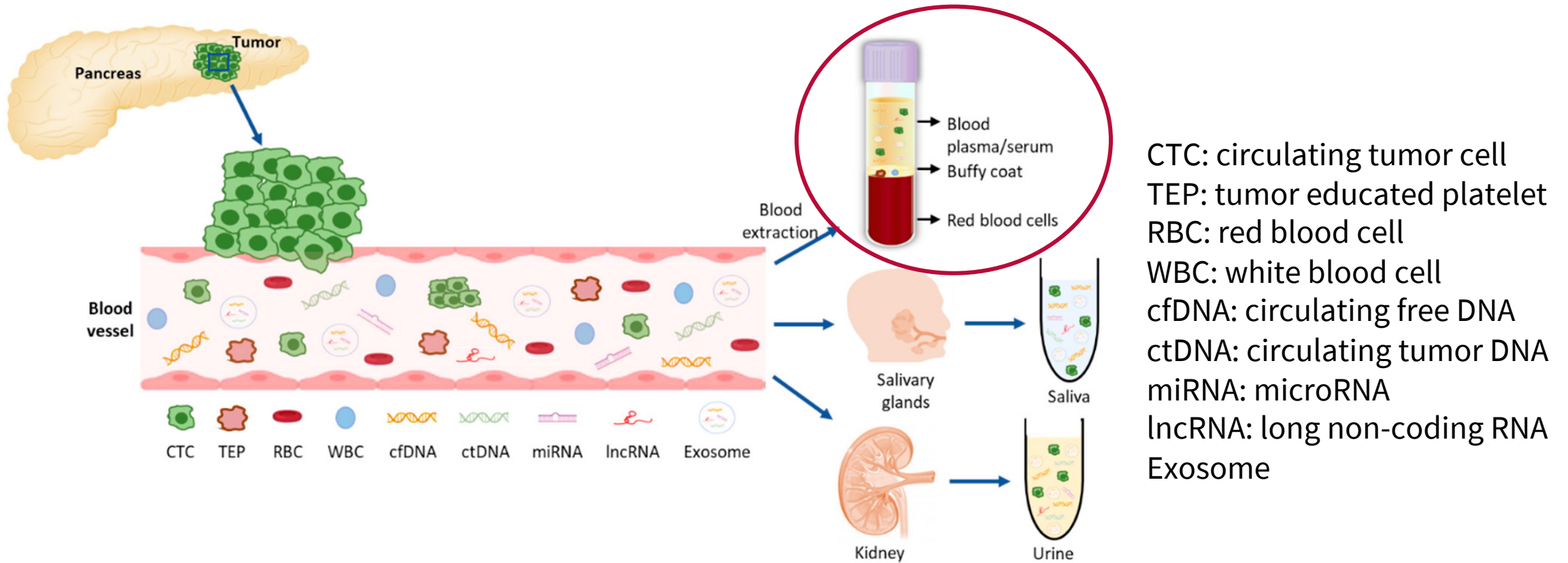
# Liquid Biopsy Applications: Prostate Cancer



Circulating tumor cells (CTCs) and extracellular vesicles (EVs), circulating cell-free tumor DNA (ctDNA) and RNA (ctRNA) all provide molecular information to characterize tumors.

Liquid biopsy is a promising minimally invasive tool for PCa management

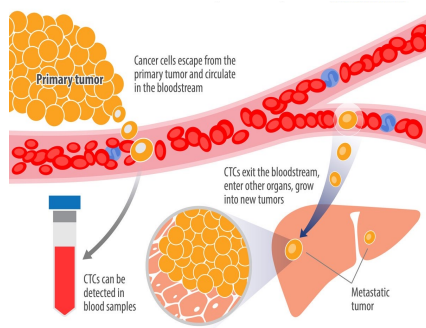
# Testing Compartment and Targets



Minimally invasive, risk-free procedure to detect the presence of genetic material from the tumor in blood and other bodily fluids, providing molecular information that reflects the global status of the disease.

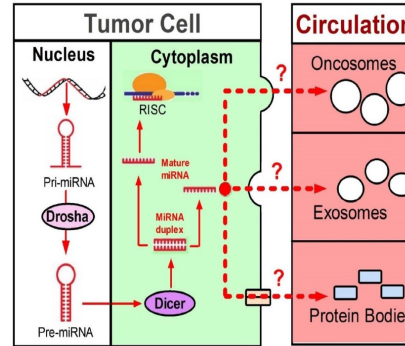
# Advantages and Disadvantages

## CTCs



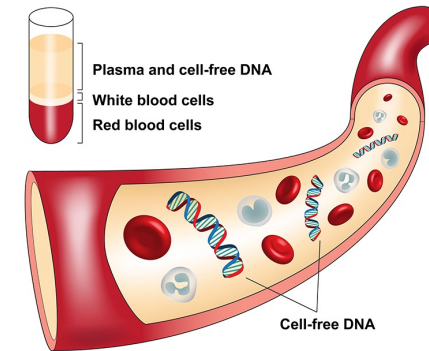
Specific  
 Single sub-population  
 Limited numbers  
 Variable purity  
 Technology evolving  
 Limited clinical utility

## miRNA/exosomes



Specific  
 Variable sensitivity  
 Contamination  
 Sample yield?  
 NGS expensive  
 Technology evolving

## cf/ctDNA



Specific  
 Variable sensitivity  
 Point mutations  
 Sample yield?  
 NGS expensive  
 Technology evolving

All applications require significant pre-analytic processing

Collection, storage and transportation

# Molecular Targeting

MUTATION

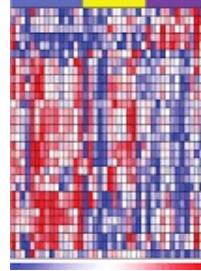


RAF Mutation

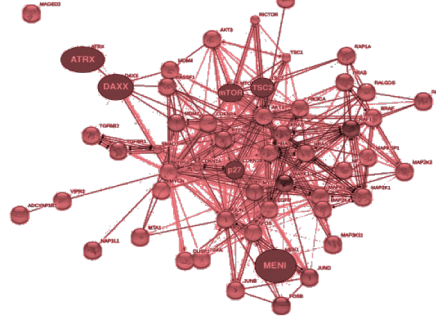
RNA



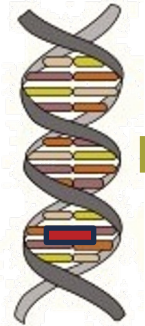
Transcriptomes



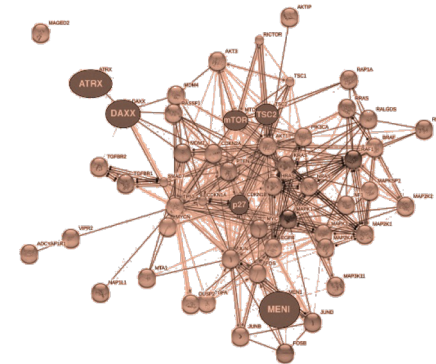
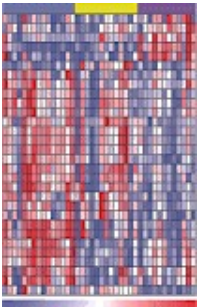
Transcriptional Networks



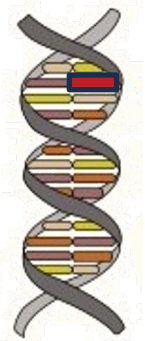
SNPs



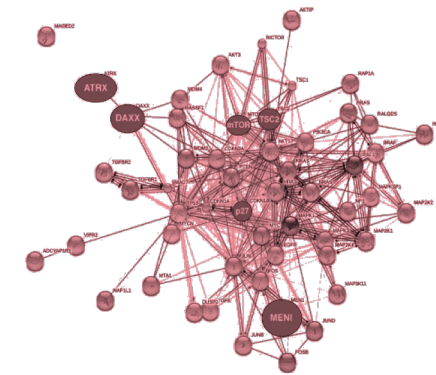
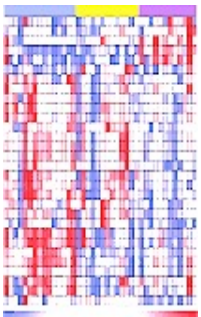
Coding SNP



EPIGENETICS



Histone Modification



mRNA is the common target

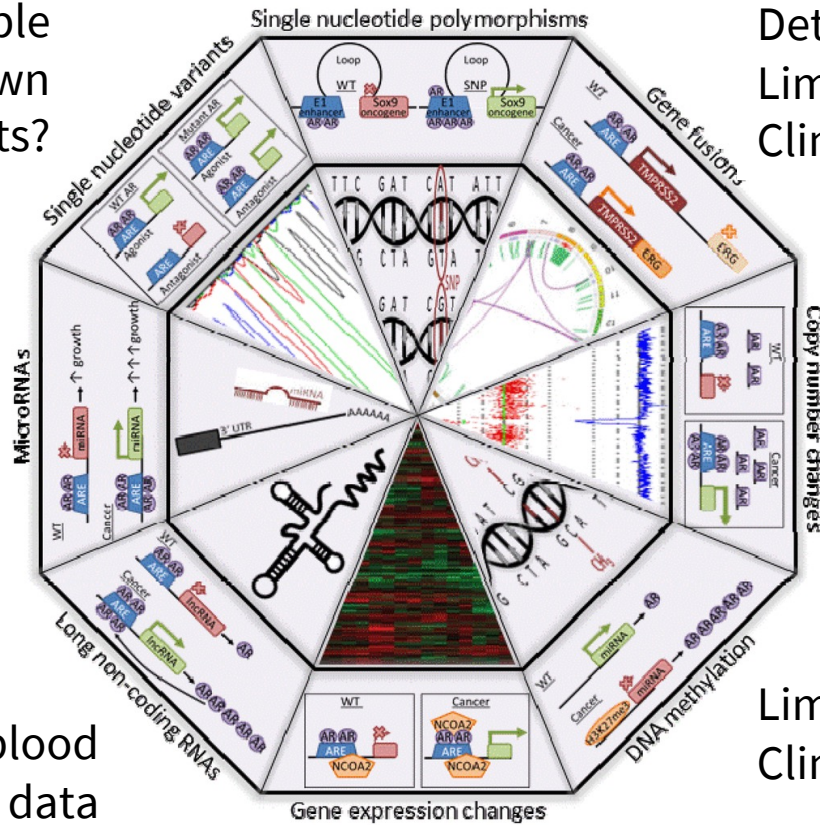
# RNA

Multiple  
Clinically useful?

Multiple  
Significance unknown  
Druggable targets?

Detectable in blood  
Informative  
Pathway activation/  
classification  
Clinically useful?

Detectable in blood  
Limited data  
Clinically useful?



Detectable in blood  
Limited data  
Clinically useful?

Limited data  
Clinically useful?

Limited data  
Clinically useful?

- **Detectable in blood**
- **Informative – cancer hallmarks**
- **Druggable targets**
- **Classification Marker of tumor behavior**

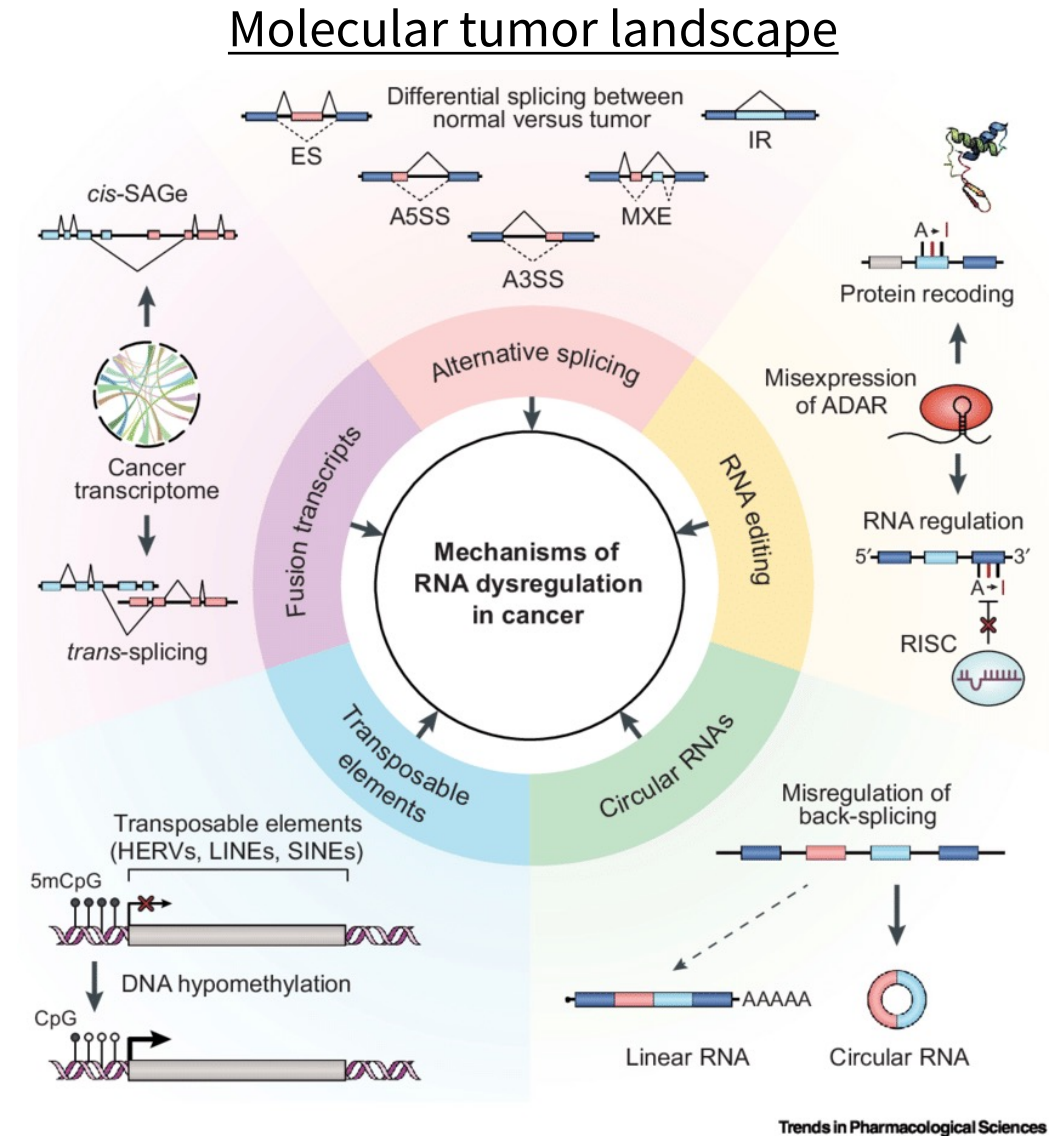
# mRNA Signatures

- Messenger RNA (mRNA) – transfers messages from DNA
- The mRNA signature of a cell - transcriptome
- Any alternation in DNA (mutations, copy number changes, epigenetic modifications) or any alterations in RNA regulation (splicing, editing) – changes the transcriptome/signature

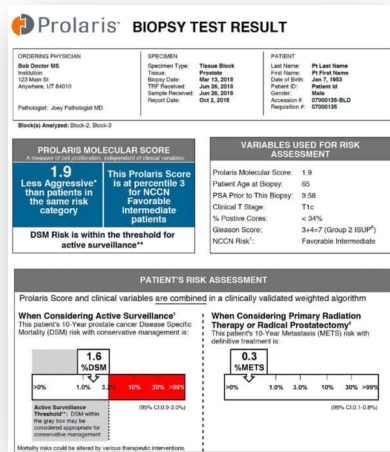
Cancer transcriptomes exhibit RNA dysregulation

mRNA is a surrogate marker of DNA status

mRNA can be used to identify tumors/tumor status

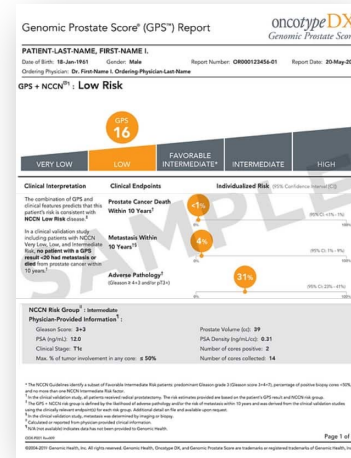


# Tissue RNA Biomarker Tools: Prostate Cancer



Progression (46)

European Urology. 2014;66(3):550-60



Tx vs. AS (21)

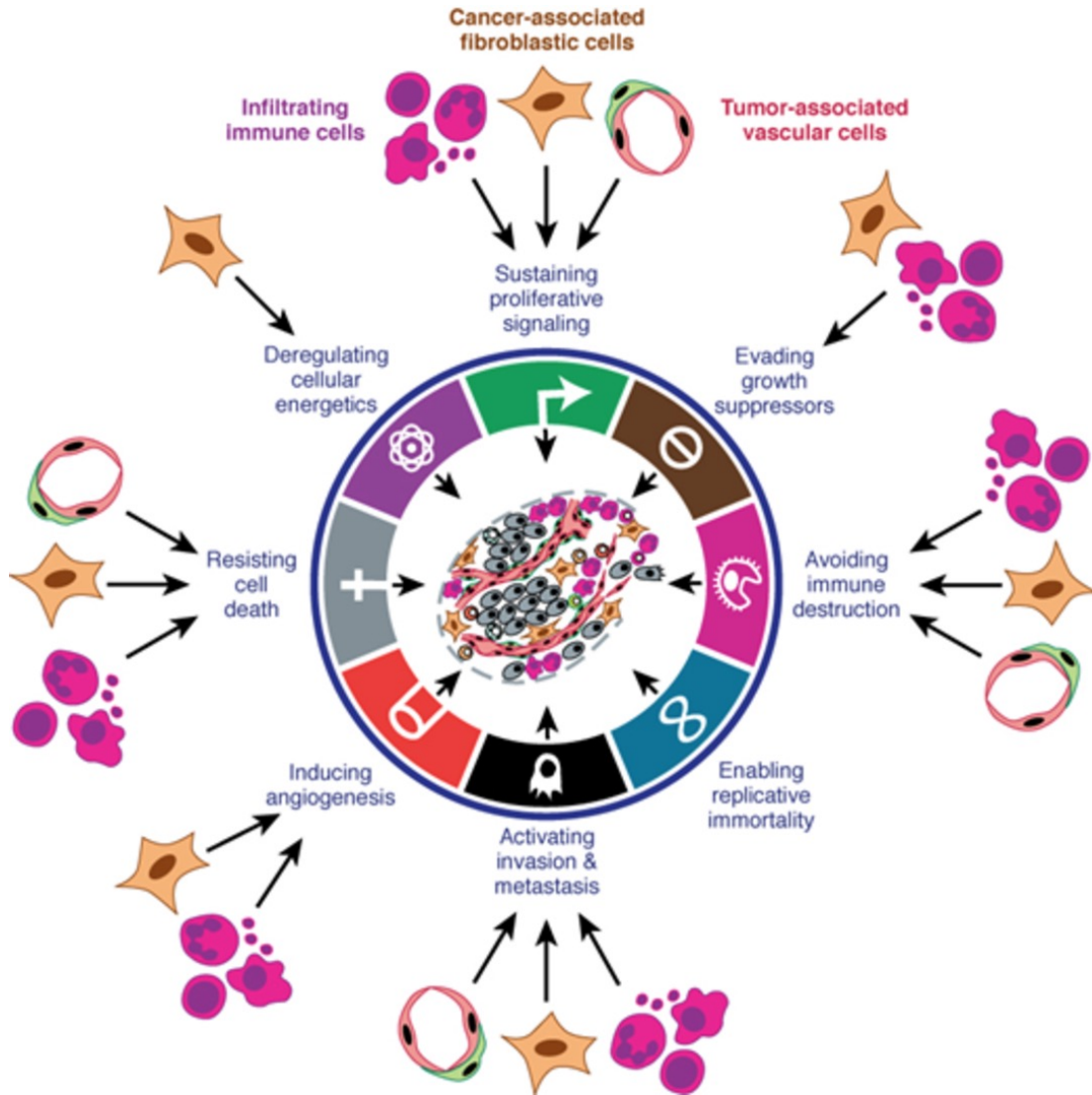
Lancet Oncol. 2011;12:245-55



Repeat Biopsy? (DNA-me)

International J Molecular sciences. 2019;20(8)

# Tumor Message is more than Tissue



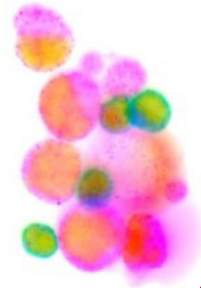
The tumor journey reflects the interaction of the genetic make-up of the lesion and the microenvironment as well as systemic responses associated with anti-neoplastic immune activity.

How do we capture the message from the tumor?

# Blood Testing Compartment

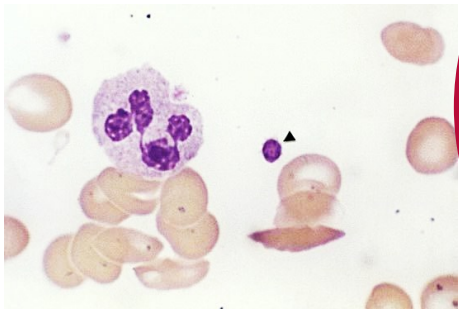
Circulating Tumor Cells

0+/10ml



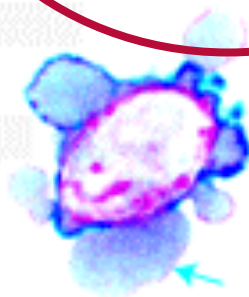
Tumor-Educated Platelets

$10^7/10\text{ml}$



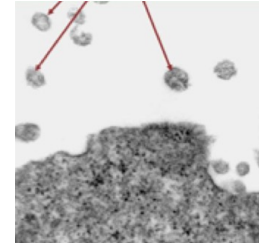
"Oncosomes"

100+ /10ml



Tumor Exosomes

$10^{10}-10^{12}/10\text{ml}$



Circulating Nucleic Acids

pg-ng/10ml



Utilize the whole blood.

# Molecular/Signature recognition

Identifying (or recognizing) the message (mRNA)

1

What is the message in the tumor cell?

How do we measure tumor mRNA?

2

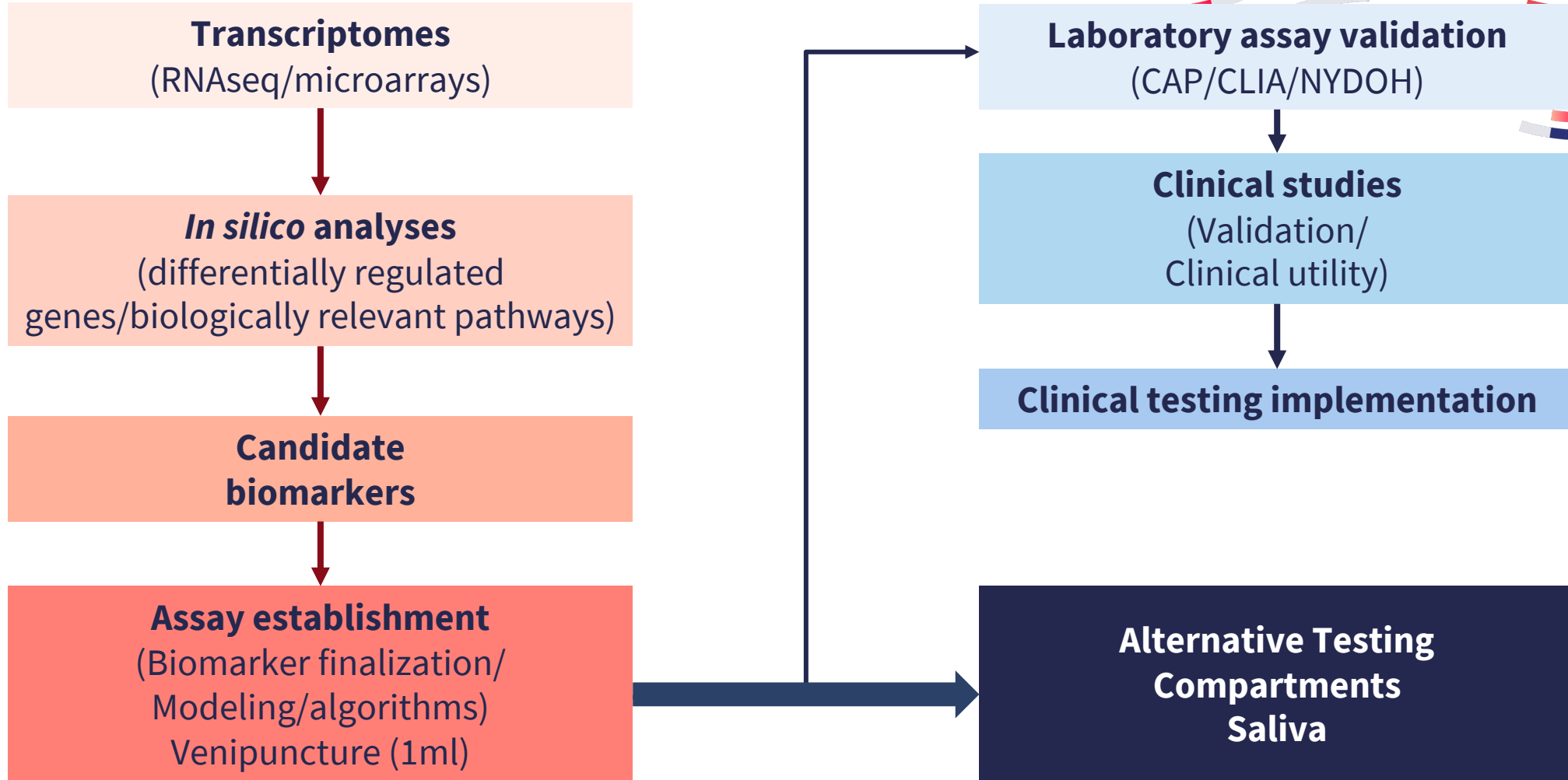
Can we measure the tumor message outside the cell?

Can the tumor message be detected in blood?

3

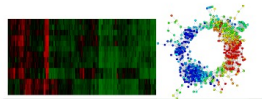
Can we build an effective assay to detect this message?

# Methodology



# Signature Recognition and Validation

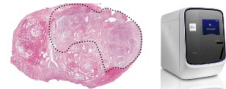
## (Prostate Cancer)



**Signature Derivation**  
 1,159 tissue transcriptomes  
 Network analysis  
 Functional enrichment  
 Differential expression



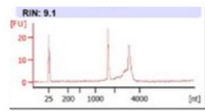
**Tumor Specificity**  
 TCGA:  $n=10,990$   
 RNAseq  
 Hierarchical clustering analysis



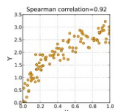
**Tissue Confirmation**  
 Surgical samples:  $n=50$   
 qPCR  
 Matched normal tissue vs. tumor samples



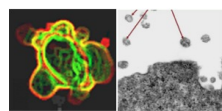
**Blood Detection**  
 65 blood samples (con, BPH, PCa)  
 Targeted RNAseq/qPCR 27 markers



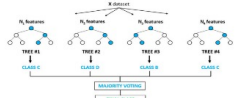
**Blood RNA Stability/Reproducibility**  
 30 controls/30 PCas  
 RIN/Bioanalyzer  
 qPCR



**Tumor:Blood mRNA Concordance**  
 50 PCas  
 Matched tumor/blood qPCR



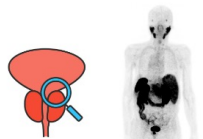
**Non-tumor Source Evaluations**  
 Whole blood  
 Platelets/Tumor blood  
 Exosomes/EVs/Immune  
 RNAseq



**Blood Algorithm Development**  
 430 age-matched subjects (control, BPH, PCA)  
 qPCR 27 markers  
 "PROSTest"



**Prospective Validation of Algorithm**  
 Age-matched controls, BPH, PCAs:  $n=303$   
 "PROSTest" vs. PSA levels/AUROC



**Surgical Validation of Algorithm**  
 Prospective,  $n=47$  radical prostatectomies  
 "PROSTest", PSA and  $^{68}\text{Ga}$ -PSMA imaging

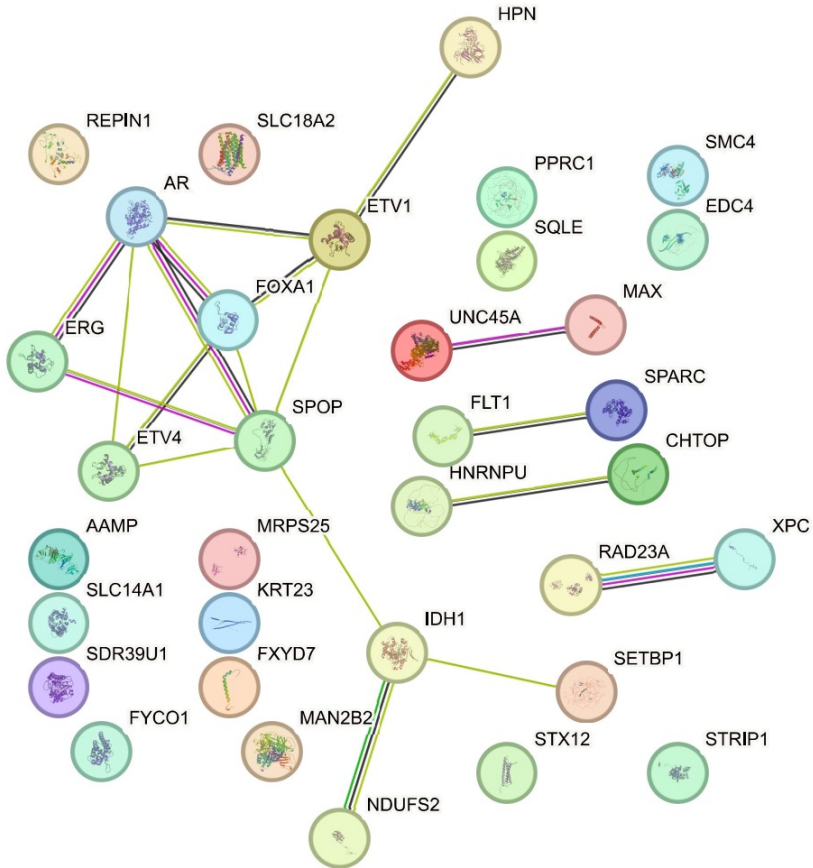
**1** Identify tumor signature

**2** Evaluate gene signature in blood testing compartment

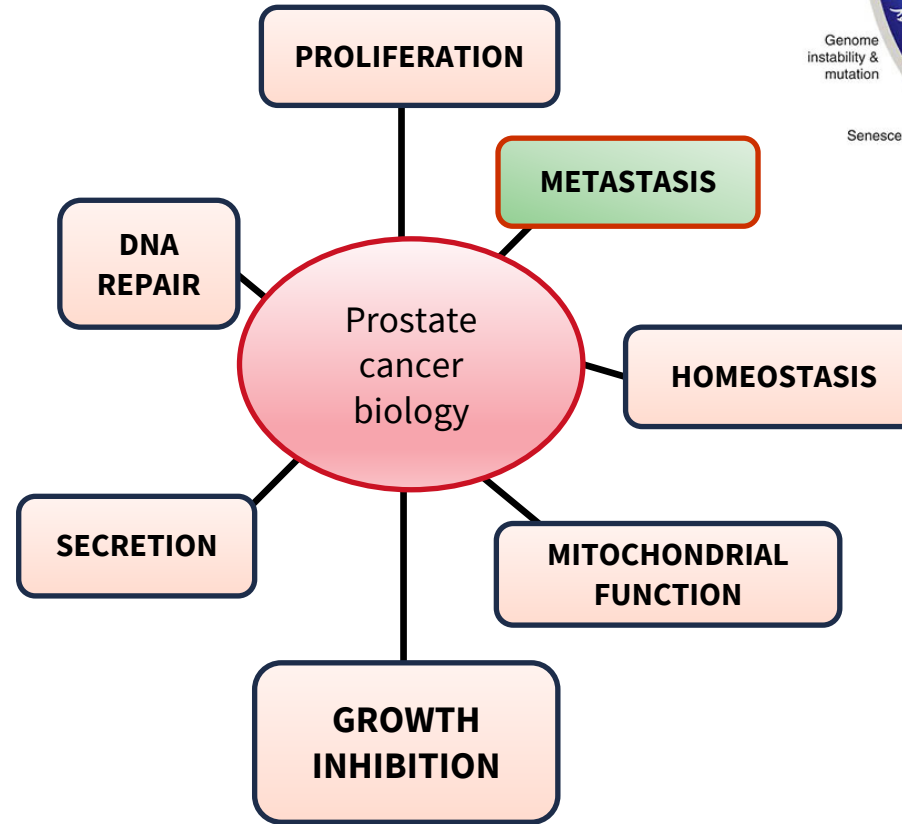
**3** Build a qPCR-assay and test algorithm



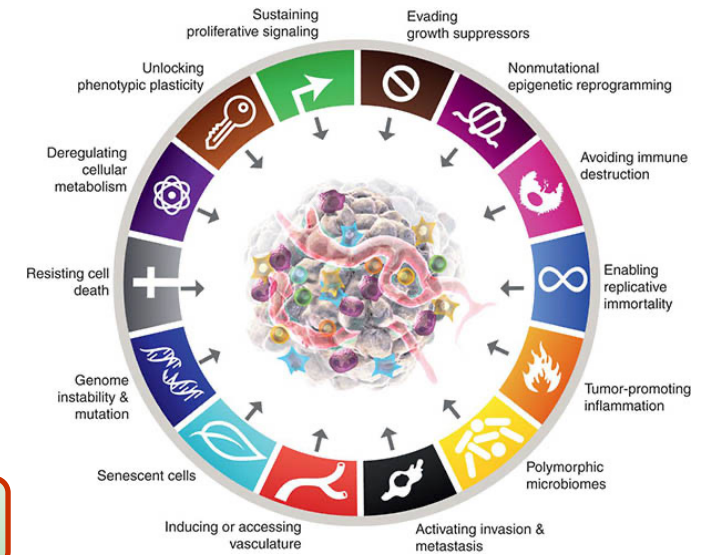
# Signature – The PROSTest



PROSTest™ INTERACTOME



PCa HALLMARKS



# Target Genes - scRNAseq

#	Symbol	Detected†	Glandular epithelial			Endothelial	Smooth muscle	Blood, immune and mesenchymal			AR regulated Gene Expression†
			Basal cell	Club cell	Glandular cell			T-cell	Macrophage	Fibroblast	
1	AAMP	Yes	43.2	36.2	35.9	28.7	33.5	30.8	21.7	27.7	Yes(-)
2	AR	Yes	178.1	80.6	499	140.4	64.5	122.4	18.3	126.7	Yes(+)
3	CHTOP	Yes	45.3	37	34.6	49.2	41.4	44	38.3	35.6	Yes(+)
4	EDC4	Yes	1.3	0.5	1.9	0.9	0.6	2.6	1.2	0	Yes(+)
5	FXYD7	Yes	0.2	0	0	0	0	3.7	0.9	0	Yes(+)
6	FYCO1	Yes	9.3	9.6	13	10.3	14.6	13.6	5	12.5	Yes(+)
7	HNRNPU	Yes	454.8	284.6	254.9	363.9	360.7	430.9	517.4	405.3	Yes(+)
8	HPN	Yes	3.8	0.6	23.4	0.3	1.8	2.3	0.7	1.3	Yes(+)
9	KRT23	Yes	77.2	41.4	2.2	19.6	3	2.6	2.9	0.7	No
10	MAN2B2	Yes	9.1	16.2	26	8.7	15.8	22	15.2	21.1	Yes(+)
11	MAX	Yes	54.4	46.8	64.4	49.8	41.4	73.8	61.2	40.3	Yes(-)
12	MRPS25	Yes	35.8	37	68.7	27.2	31	43.9	37.6	25.1	Yes(+)
13	NDUFS2	Yes	64.3	54.2	51.6	89.9	82.7	69.3	72.4	71.3	Yes(-)
14	PPRC1	Yes	11.7	6.6	5	7.5	5.5	13.1	11.3	5.9	Yes(+)
15	RAD23A	Yes	84.8	85.9	77.3	74.3	99.7	87	89.5	55.4	No
16	REPIN1	Yes	37.8	29.4	27.4	34.7	23.7	34.6	24.3	18.5	Yes(-)
17	SDR39U1	Yes	29.4	23.8	21.5	30.2	27.4	29.5	21.5	24.4	Yes(+)
18	SETBP1	Yes	27.9	6.8	20.2	19.3	45	11.5	6.5	63.4	Yes(+)
19	SLC14A1	Yes	4.6	104.5	6.3	10.9	2.4	13.3	3.1	21.8	No
20	SLC18A2	Yes	0	0	7.2	0.2	0	1.5	0.5	0	No
21	SMC4	Yes	39.4	14.3	7.8	34.7	24.3	27.7	32.5	17.2	Yes(-)
22	SPARC	Yes	59.9	4.7	0	370.8	225	8.5	5.5	233.7	No
23	SQLE	Yes	15.6	26.1	43.3	10.6	9.1	12.4	22.4	7.3	No
24	STRIP1	Yes	7.3	6.1	10.7	7.5	2.4	7.5	7	5.3	No
25	STX12	Yes	60.1	33.6	103.1	45.3	32.8	70.8	54.8	29	Yes(-)
26	UNC45A	Yes	22.1	31.7	28.1	30.2	43.8	24.6	17.8	17.8	No
27	XPC	Yes	60.6	40.9	37.8	45.3	35.9	59.9	36.4	43.6	Yes(-)

\*mRNA expression detected in single cells

†Protein Atlas single cell evaluation <https://www.proteinatlas.org/gene/single+cell+type/prostate> nTPM 3

†AR = androgen receptor signaling. Gene expression is regulated by androgen receptor activation (from RNAseq data-evaluation of differential expression following 24 hr incubation with either 10nM dihydrotestosterone [AR- agonist] or 10µM bicalutamine [AR- antagonist] in LNCaP cells. From Shah et al. Oncogene 2020; 39:6172-6189, Supplemental dataset 1) 4. No: no experimental evidence; Yes(+): activated; Yes(-): repressed.

# Immune Cells

#	Symbol	Whole Blood (TPM)*	PBMCs (nTPM)‡	Single cell flow-sorted (nTPM)‡	Role in Hematopoiesis (DMAP)† **	Immune cell specificity and cell distribution‡
1	AAMP	0.05	Monocyte (60) B-cell (60)	ncMonocyte (212) iMonocyte (209) Myeloid DC (192)	Erythrocyte	Low immune cell specificity
2	AR	0.02	DC (40) Platelets (38)	ND	None	Not detected in immune cells
3	CHTOP	21.0	T-cell (80) B-cell (74)	Basophil (85)	B-cell CD4/CD8 Monocyte Neutrophil	Low immune cell specificity
4	EDC4	17.5	NK cell (0.9)	ncMonocyte (11) T-reg cell (11)	B-cell CD4	Low immune cell specificity
5	FXYD7	2.1	T-cell (16)	NK cell (52)	None	Immune cell enhanced (NK-cell)
6	FYCO1	2.3	T-cell (14) DC (14)	MAIT T-cell (2.6) GdT-cell (2)	None	Low immune cell specificity
7	HNRNPU	33.7	Macrophage (378) DC (317)	pDC (31.4) Naive CD4 (25.5)	B-cell CD4/CD8 Erythrocyte Neutrophil	Low immune cell specificity
8	HPN	0.50	ND	ND	None	Not detected in immune cells
9	KRT23	0.50	Macrophage (3)	Neutrophil (0.9)	None	Not detected in immune cells
10	MAN2B2	12	Monocyte (18) Macrophage (17)	ncMonocyte (18) Basophil (16)	B-cell CD4/CD8 Monocyte Eosinophil Neutrophil	Low immune cell specificity
11	MAX	56.8	Platelet (2,007)	Basophil (670) Eosinophil (542)	B-cell CD4/CD8 Neutrophil	Low immune cell specificity
12	MRPS25	5.5	T-cell (60) Monocyte (44)	Memory B-cell (65)	None	Low immune cell specificity
13	NDUFS2	29.8	T-cell (73) Monocyte (59)	Myeloid DC (218) iMonocyte (193)	Erythrocyte	Low immune cell specificity
14	PPRC1	3.7	NK cell (9) T-cell (8)	NK cell (4.1)	None	Low immune cell specificity
15	RAD23A	73.1	T-cell (122) B-cell (120)	ncMonocyte (106) iMonocyte (102)	Erythrocyte	Low immune cell specificity
16	REPIN1	17.7	DC (97)	pDC (7.8)	B-cell Neutrophil	Immune cell enhanced (pDC)

#	Symbol	Whole Blood (TPM)*	PBMCs (nTPM)‡	Single cell flow-sorted (nTPM)‡	Role in Hematopoiesis (DMAP)† **	Immune cell specificity and cell distribution‡
17	SDR39U1	9.8	Platelet (48) T-cell (42)	pDC (62) NK cell (60)	None	Low immune specificity
18	SETBP1	0.5	DC (47) B-cell (35)	Naive B-cell (2.9) pDC (2.7)	Clonal Hematopoiesis	Immune cell enhanced (B/pDC)
19	SLC14A1	3.5	T-cell (19) NK cell (11)	T-reg (4.1)	None	Immune cell enhanced (T-reg)
20	SLC18A2	0.2	Platelet (48) DC (14)	Basophil (60)	None	Immune cell enriched (basophil)
21	SMC4	2.0	T-cell (87)	Basophil (75)	None	Immune cell enhanced (basophil)
22	SPARC	33.5	Platelet (2,130)	Neutrophil (25) Basophil (10)	Erythrocyte Neutrophil B lymphopoiesis	Group enhanced (neutrophil, basophil, classical monocyte)
23	SQLE	2.0	B-cell (29)	Naive B-cell (17) Memory B-cell (16.6)	None	Low immune cell specificity
24	STRIP1/ FAM40A	10.6	T-cell (18) B-cell (18)	NK cell (16) Memory CD8 (16)	NA	Low immune cell specificity
25	STX12	13.7	Macrophage (61) Monocyte (51)	ncMonocyte (41) iMonocyte (40)	B-cell Monocyte Neutrophil	Low immune cell specificity
26	UNC45A	18.6	Platelet (57)	GdT-cell (11.8) ncMonocyte (11.7) Basophil (11.5)	B-cell	Low immune cell specificity
27	XPC	13.2	T-cell (61) NK cell (57)	Neutrophil (74)	CD4/CD8	Low immune cell specificity

\*From GTex ([www.gtex.portal.org/home/gene](http://www.gtex.portal.org/home/gene))<sup>5,6</sup> (n=755 whole blood samples, median TPM)

‡From the HPA ([www.proteinatlas.org/](http://www.proteinatlas.org/)) (n=109 blood samples, nTPM)<sup>7</sup>

†From BloodSpot (Haemopedia) DMAP<sup>8,9</sup> – normal human Hematopoiesis: <https://servers.binf.ku.dk/bloodspot/?gene=>. Gene signatures are derived from DMAP data ([www.broadinstitute.org/dmap/home](http://www.broadinstitute.org/dmap/home))

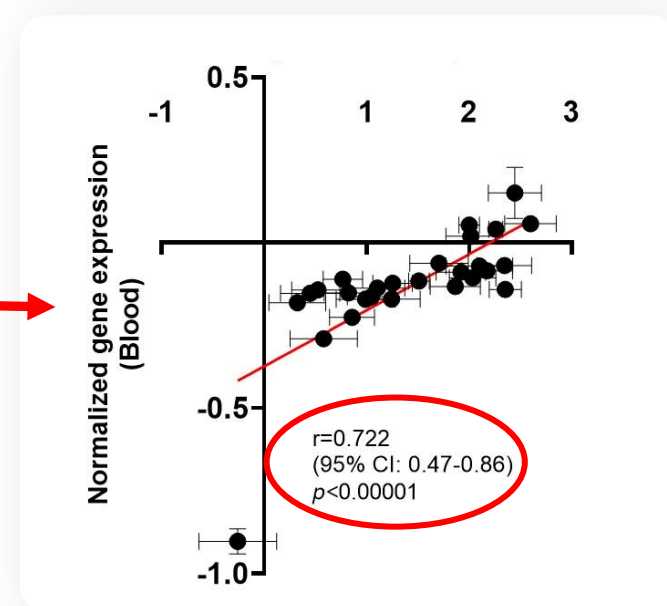
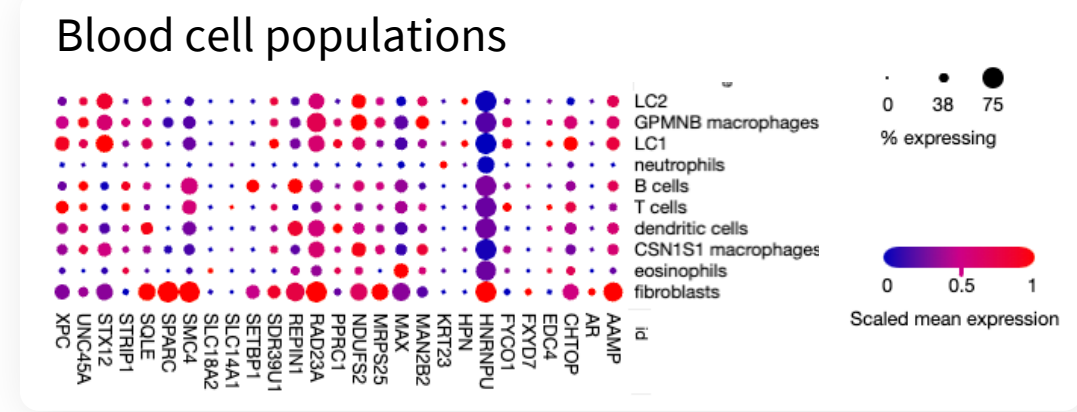
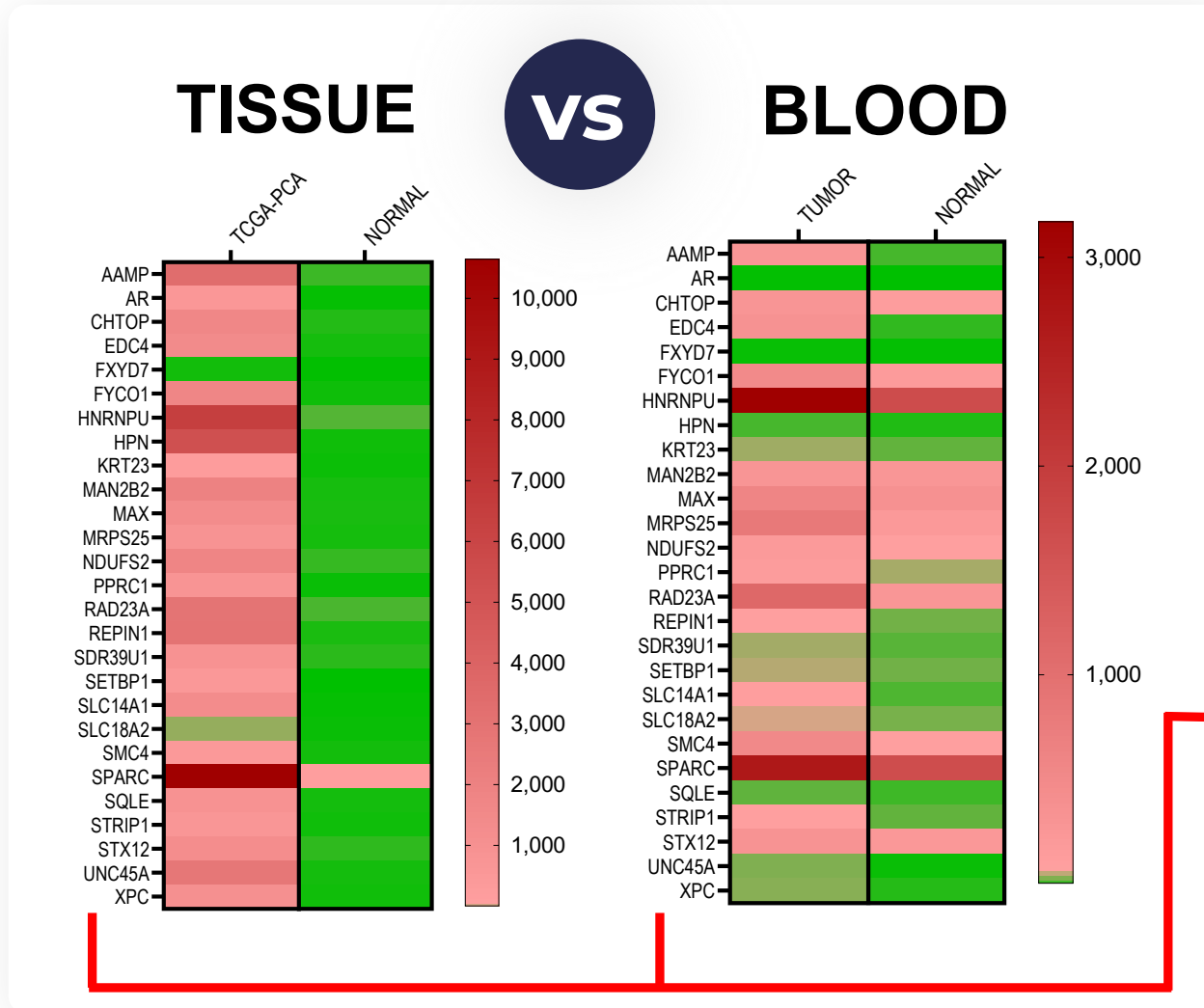
\*\*PubMed search (gene name/Hematopoiesis, 9/2023)

Low immune cell specificity = detected in all immune cells at low levels

DC = dendritic cell, GdT = gamma-delta T-cell, iMonocyte = intermediate monocyte, MAIT = mucosal associated invariant T cell, NA = not annotated, ncMonocyte = non-classical monocyte, ND = not detected, pDC = plasmacytoid dendritic cell

# Molecular Recognition

Is it tumor-related or is the message blood-derived?



**Tumor**  
Tissue: Blood  
Correlation

# Molecular Recognition – Algorithms

## How accurate are blood-based mRNA signatures?

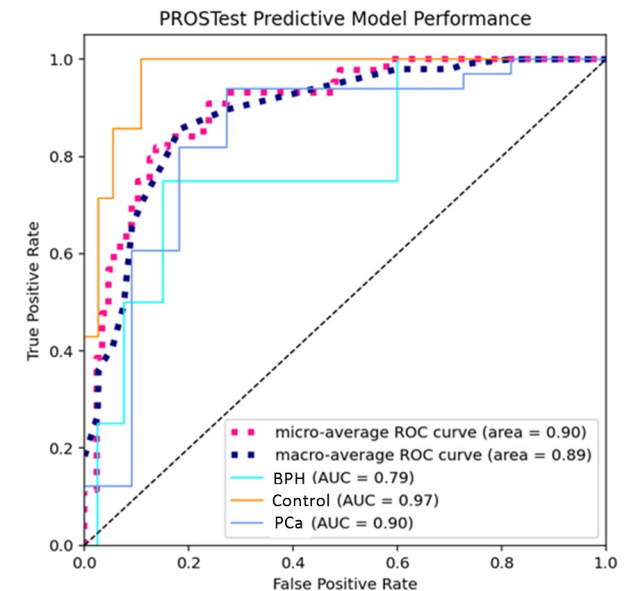
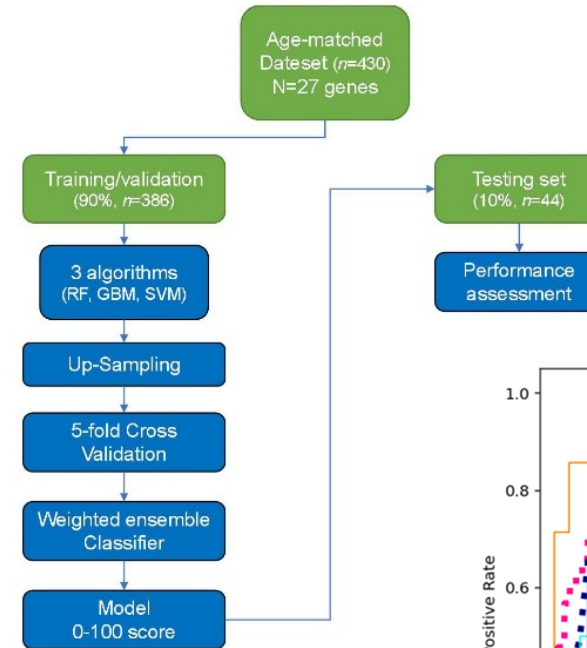
Classifiers function to identify patterns of gene expression and “learn” to assign samples to a specific class (e.g., cancer vs. control).

Hyperparameter optimization (HPO) and feature selection (FS) improve the predictive performance of algorithms

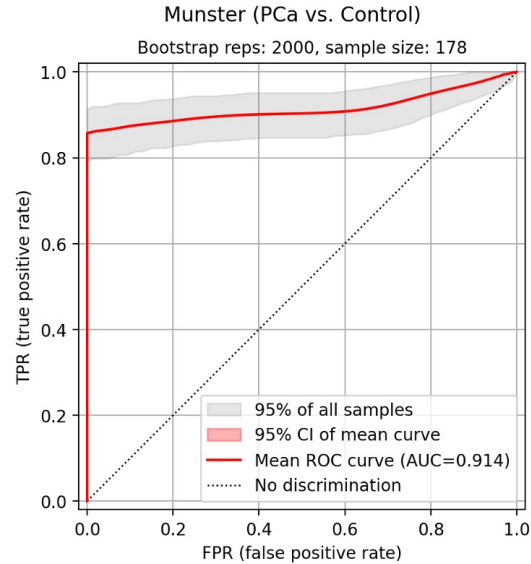
Ensemble-based algorithms consistently outperform other types of classification algorithms



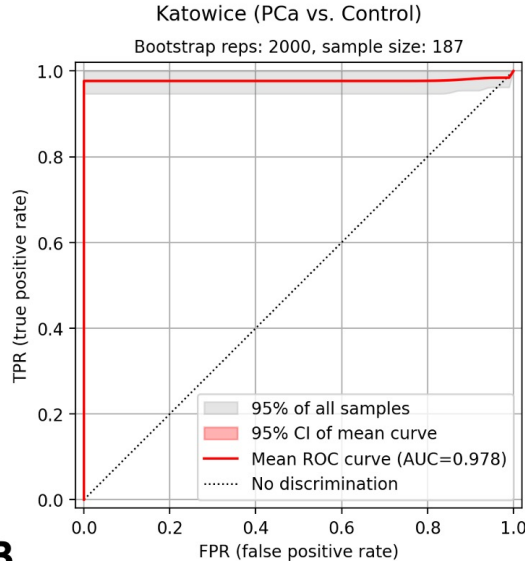
Wren algorithms utilize HPO and FS



# Algorithm Testing

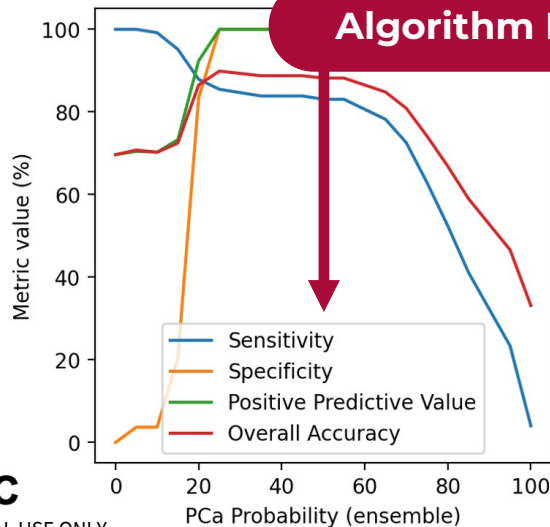


**A**

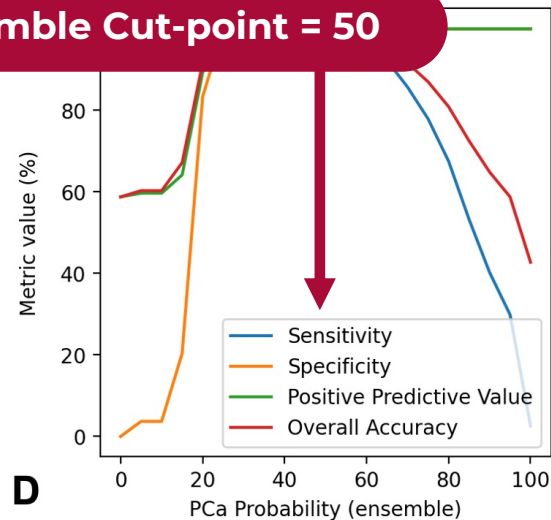


**B**

- A. Receiver operator curves (ROC) for differentiating age-matched PCa and Controls (Cohort I:  $n=178$ )
- B. (ROC) for differentiating age-matched PCa and Controls (Cohort II:  $n=187$ )



**C**



**D**

- C. Line plots of PPV, sensitivity, specificity (Cohort I)
- D. Line plots of PPV, sensitivity, specificity (Cohort II)

# Algorithm Performance Metrics\*

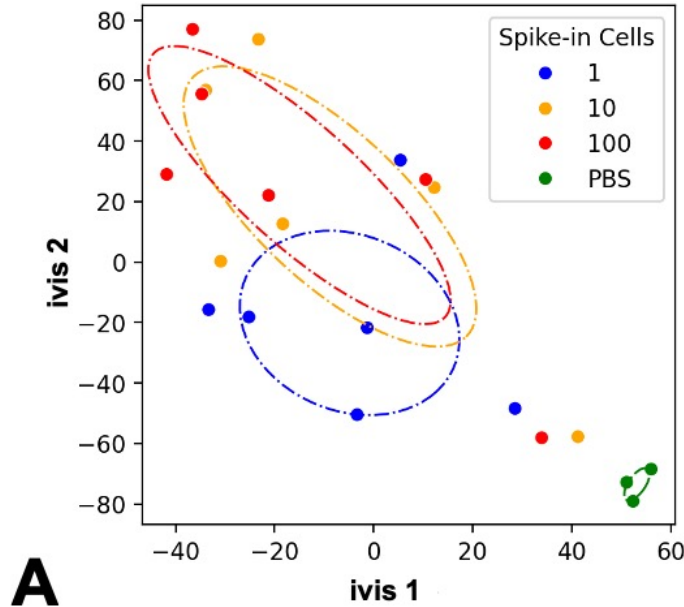
Dataset	Class	AUROC	Sensitivity	Specificity	PPV
<b>Munster</b> <b>(n=178)</b>	PCa vs. Control	0.91 (0.913-0.915)	83% (80%-87%)	100% (96%-100%)	100% (95%-100%)
<b>Katowice</b> <b>(n=187)</b>	PCa vs. Control	0.98 (0.977-0.979)	94% (92%-96%)	100% (96%-100%)	100% (96%-100%)

95% Confidence Intervals are shown in parentheses. PPV=Positive Predictive Model.



\*Using the Algorithm Ensemble Cut-point = 50 (scale 0-100)

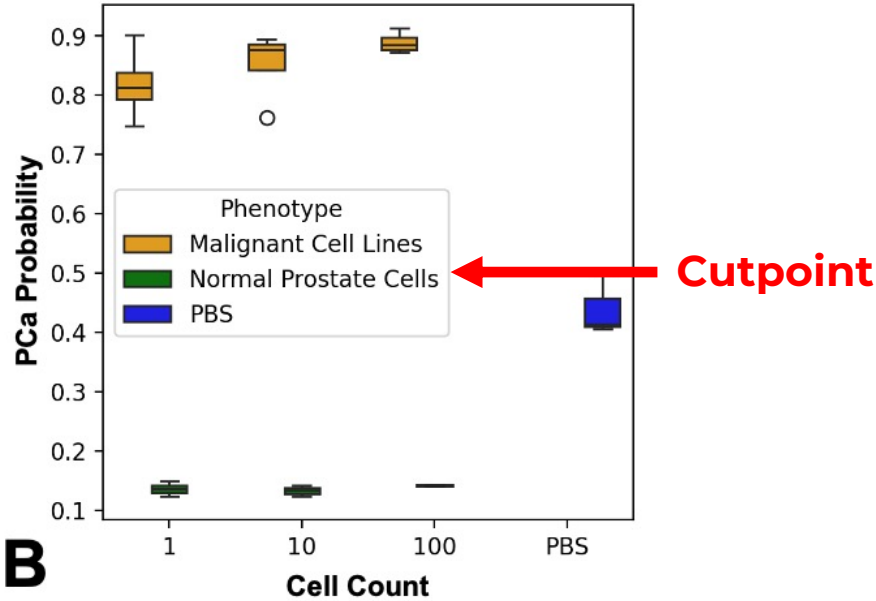
# Blood Spike-in studies



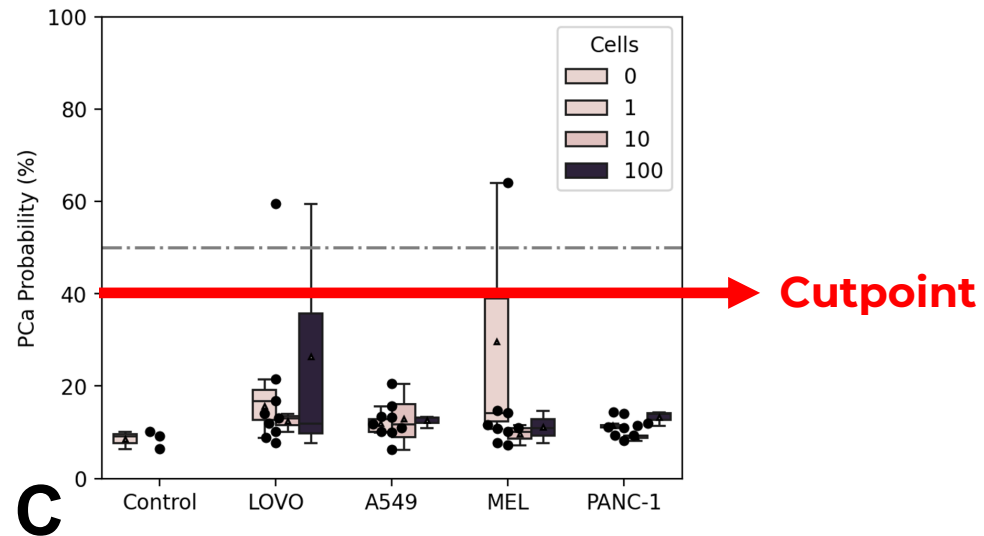
**A**

**Normal prostate:**  
PWRE-1, RWPE-1

**Malignant PCa:**  
DU145, LnVAP, MDA PCa2b, PC-3



**B**



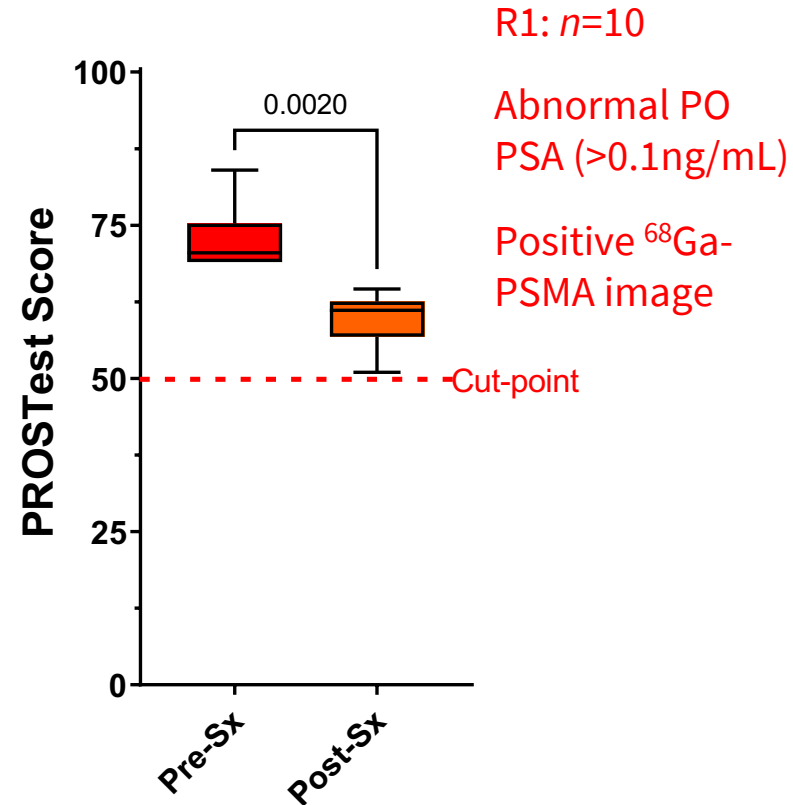
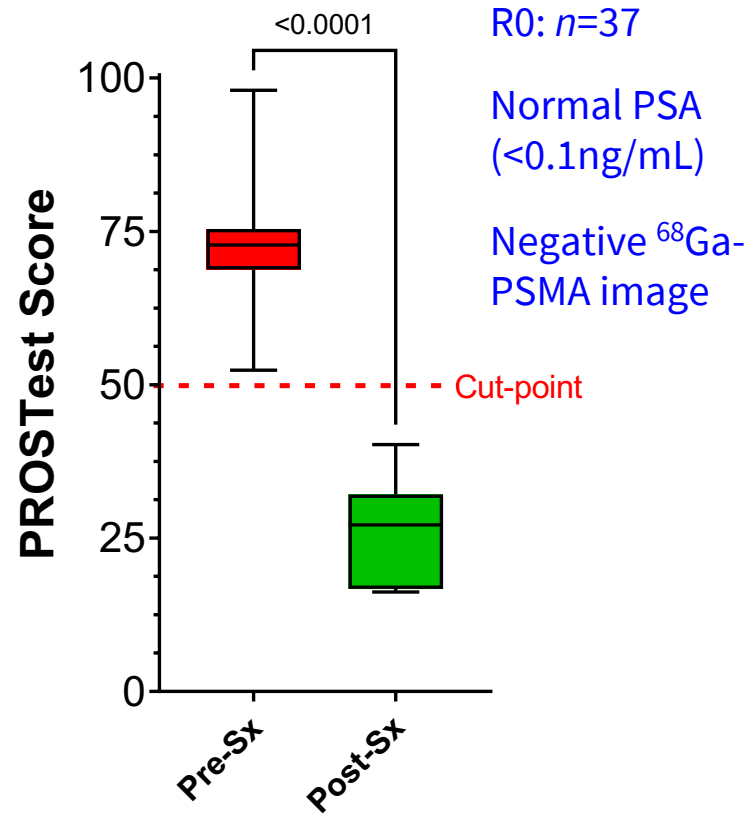
**C**

- A. Scatterplot showing two-dimensional twin neural network (Ivis) embedding of the model feature values in spike-in experiments.
- B. Boxplots of spike-in blood algorithm scores – prostate cancer cell lines ( $n=6$ )
- C. Boxplots of spike-in blood scores – non-prostate cancer cell lines ( $n=4$ )

LOVO = colon  
A549 = lung  
MEL = A375/melanoma  
PANC-1 = pancreas

# Surgery and Blood Gene expression

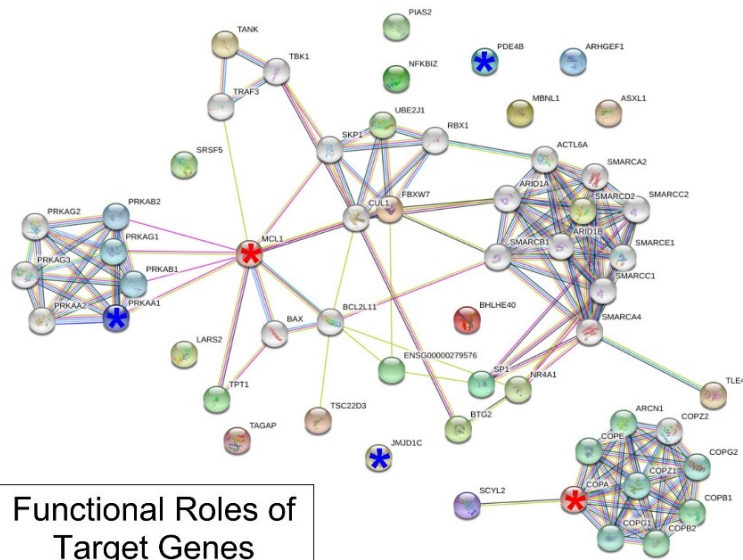
(Radical Prostatectomy cohort)



Complete tumor removal removes source of the “PROSTest” signal

**!** Blood gene expression “PROSTest” is derived from tumor tissue

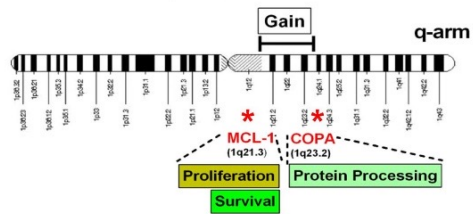
# Multiple Myeloma



Functional Roles of Target Genes

Apoptosis	Immune Response	Phenotype	Protein Processing	Proliferation	RNA Processing	Survival
BTG2 NFKBIZ	LARS2 NFKBIZ NR4A1 TAGAP TANK TLE4 TSC22D3	NR4A1 SCYL2 SMARCD2 TLE4	COPA1 LARS2 SCYL2 UBE2J1	GNA13 JMJD1C MALAT1 MCL-1 PDE4B	MBNL1 SRSF5	MCL-1 NFKBIZ PDE4B PRKAA1 TSC22D3 UBE2J1

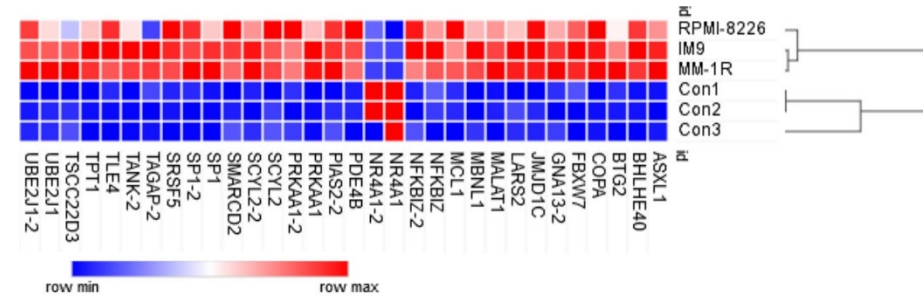
Chromosome 1



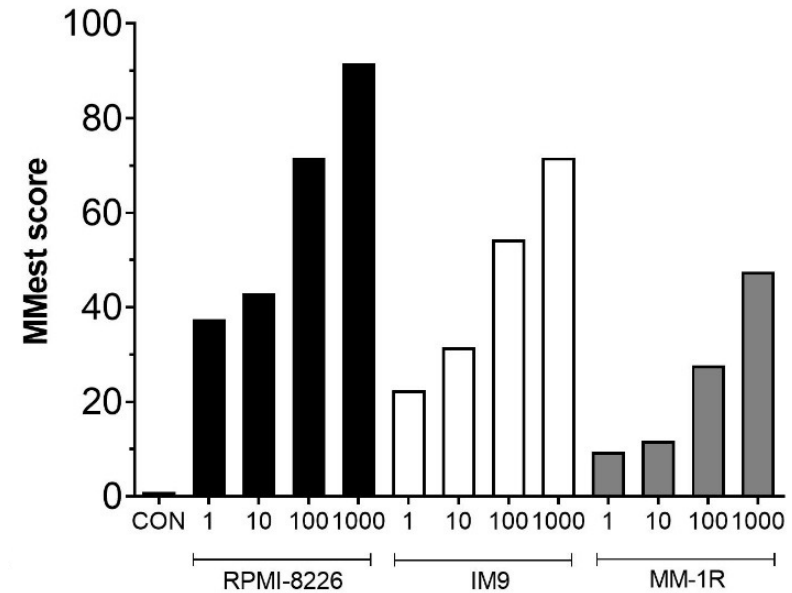
\* Potential Drug Targets  
PRKAA1 (AMP)  
PDE4B (AKT/mTOR)  
JMJD1C (anti-leukemics)

MALAT  
Oncogene  
(long non-coding RNA)

Hierarchical clustering

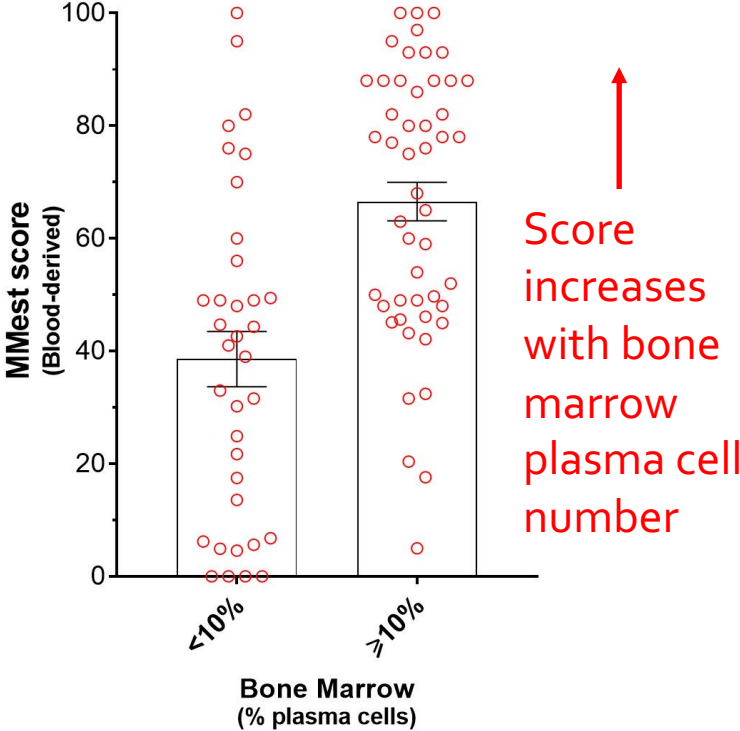


Blood spike-in study

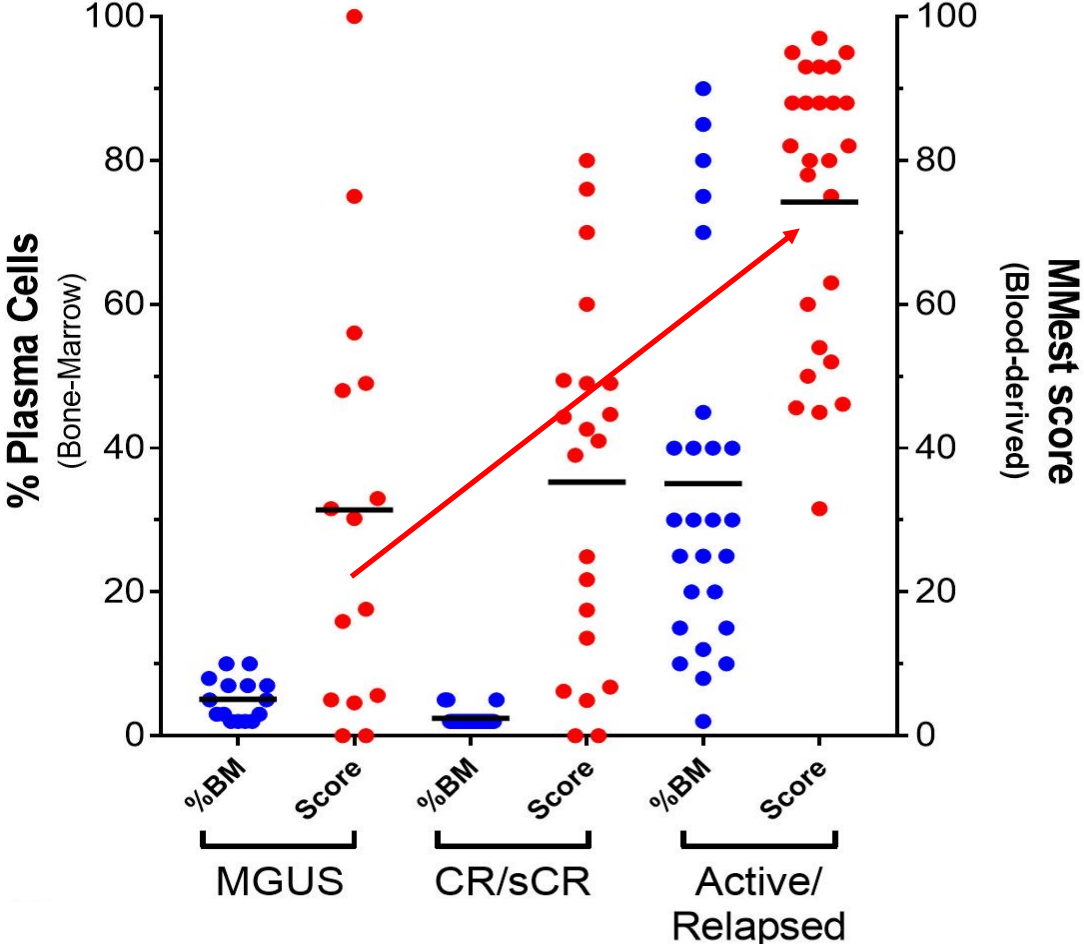


# Multiple Myeloma

25-gene blood algorithm score (MMest)



Pearson correlation:  $r=0.29$ ,  $p<0.006$ ,  $n=84$



Horizontal black lines = mean levels  
CR = complete remission

# Neuroendocrine Tumor – NETest®



## A meta-analysis of the accuracy of a neuroendocrine tumor mRNA genomic biomarker (NETest) in blood

K. Öberg<sup>1\*</sup>, A. Califano<sup>2</sup>, J. R. Strosberg<sup>3</sup>, S. Ma<sup>4</sup>, U. Pape<sup>5</sup>, L. Bodei<sup>6</sup>, G. Kaltsas<sup>7</sup>, C. Toupanakis<sup>8</sup>, J. R. Goldenring<sup>9</sup>, A. Frilling<sup>10</sup> & S. Paulson<sup>11</sup>

<sup>1</sup>Department of Endocrine Oncology, University Hospital, Uppsala, Sweden; <sup>2</sup>Department of Systems Biology, Columbia University, New York; <sup>3</sup>Department of Gastrointestinal Oncology, Moffitt Cancer Center, Tampa; <sup>4</sup>Department of Biostatistics, Yale University, New Haven, USA; <sup>5</sup>Department of Internal Medicine and Gastroenterology, Asklepios Kliniken Hamburg, Germany; <sup>6</sup>Department of Radiology, Memorial Sloan Kettering Cancer Center, New York, USA; <sup>7</sup>Division of Endocrinology, Department of Pathophysiology, University of Athens, Athens, Greece; <sup>8</sup>Department of Gastroenterology, University College of London, London, UK; <sup>9</sup>Section of Surgical Sciences, Vanderbilt University Medical Center, Nashville, USA; <sup>10</sup>Department of Endocrine Surgery, Imperial College London, London, UK; <sup>11</sup>Division of Medical Oncology, Baylor Charles A Sammons Cancer Center, Dallas, USA

Annals Oncology 2020; 31: 202-212

### In vitro diagnostic (IVD)

**Accuracy: 96%**

Sensitivity: 94.4%

Specificity: 98.7%

### Natural History

**Accuracy: 94%**

Sensitivity: 99.6%

Specificity: 99.4%

### Disease Status

**Accuracy: 86%**

Sensitivity: 83.1%

Specificity: 89.4%

### Monitor Therapy

**Accuracy: 97%**

Sensitivity: 90.1%

Specificity: 99.7%

The NETest is an accurate biomarker suitable for clinical use in NET disease management.

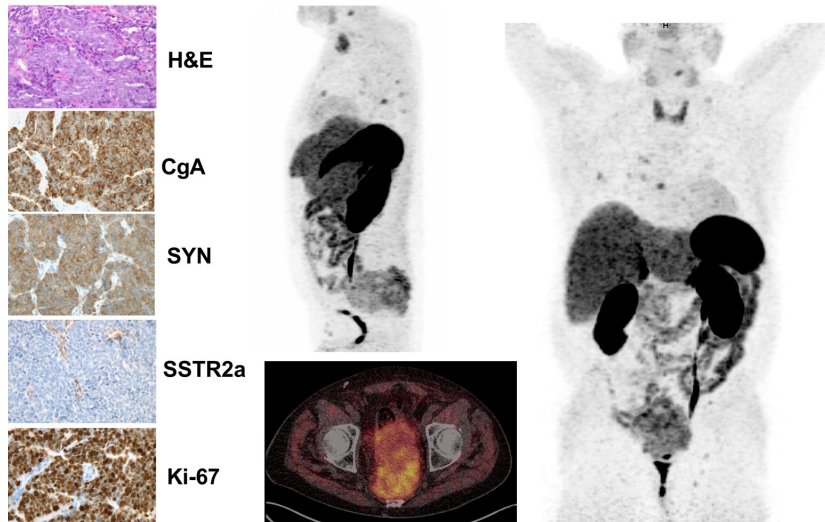
The meta-analysis supports the utility of the NETest as an IVD to establish a diagnosis and monitor therapeutic efficacy.

The use of this as a biomarker provides information relevant to NET management consistent with observations regarding utility of liquid biopsies in other oncological disciplines.

# Blood-based detection of Neuroendocrine Differentiation

## NETest<sup>®</sup>

### Tumor Neuroendocrine-like Differentiation (Neld/Ned)

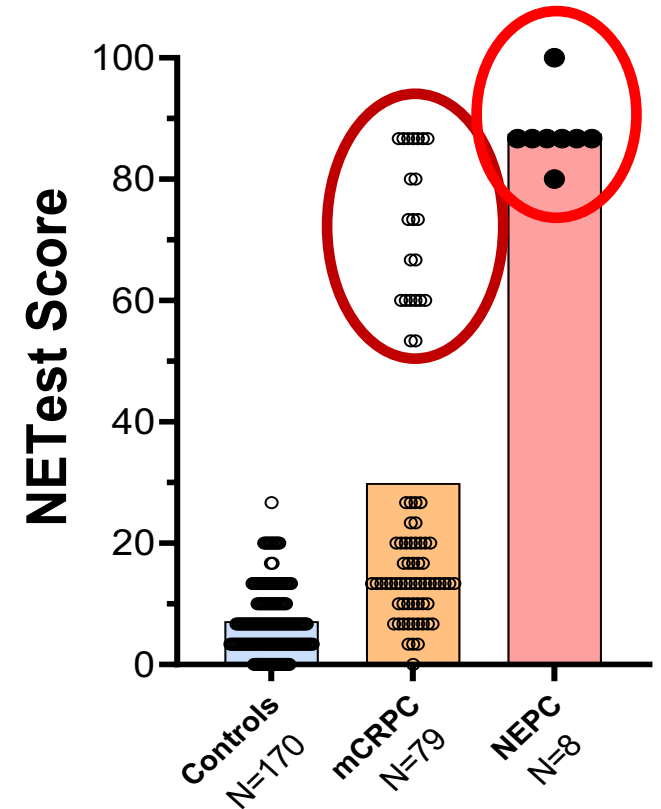


Prostate (PRAD)

Lung (LUAD)

Colon (COAD)

Early detection  
Therapy Failure  
(e.g., ADT, targeted therapies)



# NETest – A Commercial Success

Over 9,000 NETests used in numerous global studies with leading US and international academic and clinical institutions.

## Successful and profitable NETest™ Pharma collaborations



Phase 1 Clinical trial of XmAb®18087:  
NETest biomarker for treatment efficacy  
- XmAb18087 engages the immune system against tumors by binding to somatostatin receptor 2 (SSTR2) and CD3.



Von Hippel Lindau germline tumors:  
NETest: Diagnostic biomarker  
  
NETest: Diagnostic biomarker:  
Parangliomas and Pheochromocytomas



NETest: Clinical testing  
Diagnosis, monitoring, surgery, PRRT  
**Beijing, China**, Peking Medical University  
Precision Medicine Oncology



Phase 2 Open label study of 212Pb-AlphaMedix targeted alpha therapy:  
NETest biomarker for treatment efficacy  
- Somatostatin receptor 2 (SSTR2) radiotherapy targeting metastatic NETs.



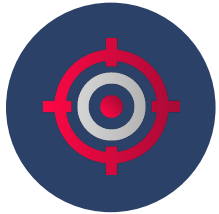
NETest: Clinical testing  
PRRT prediction, monitoring  
Precision Medicine Oncology USA



NETest: Clinical testing  
Diagnosis, monitoring, PRRT  
**Mexico**, Precision Medicine  
Oncology

# Our Approach: Assay Development

mRNA based disease biomarkers and patient clinical data are the basis of our diagnostic approach



## Discover mRNA-based Biomarkers

Identify genes with RNA expression levels significantly different from controls or between disease states



## Incorporate Patient Data

Utilizing ML algorithms to create hyperparametric gene expression data from patient demographics allows us to go beyond simple diagnostics



## Develop Diagnostic Approach

Diagnostic, prognostic, and monitoring outputs provide insight on patient journey using either blood or saliva samples

# Appendix



# Wren Laboratories: Founded 2012



## Platform

A molecular diagnostics company that develops mRNA-based biomarkers for assessment with a suite of AI-enabled multianalyte algorithms to identify and characterize a range of cancers and other diseases



## Proven Success

Innovator of molecular signatures for cancer diagnosis and management. Our liquid biopsy-based diagnostics use molecular science to uncover precise information about cancer in real-time, enhancing pathology and imaging.



## Available Diagnostics

NETest (2014)  
PPQ (2022)  
SARS-CoV-2 (2021)

**Coming soon :**  
PROSTest 2024



## Credentials

CLIA Certified  
CAP Accredited  
CT Clinical Lab License CL-0996  
New York Dept of Health  
Clinical Lab Permit PFI: 9138  
Florida License 800027786,  
certificate 101250  
PA Laboratory ID: 34347  
California Lab ID:  
COS00800634

# The Team

The Wren Laboratories team is comprised of individuals with expertise ranging from medical, clinical, research and commercial diagnostic execution.



**Irvin M. Modlin**

Founder, Clinical &  
Scientific Consultant  
MD, Ph.D., DSc



**Daniel Buck**

Managing Director  
MBA, MPH



**Mark Kidd**

Scientific & Laboratory Director  
Ph.D., DABCC



**Ignat Drozdov**

Scientific & Clinical AI Consultant  
MD, Ph.D.



**Andi Neugarten**

Corporate & Business  
Development  
B, Acc, CA.SA



**Mark Harman**

Medical & Clinical Affairs  
MD, MBA, MS ,BCMAS



**Srinivas Koduru**

Transcriptomics  
PhD



**Tracy Auster**

Laboratory Manager  
ASCP, AGT

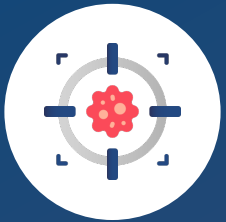


**EJ Vongher**

Marketing Director

# Wren Laboratories

Molecular-based tools that accurately capture tumor pathobiology



Diagnosis



Treatment  
Prediction



Monitoring



Disease  
Prognosis



# WREN Test Pipeline: A Development Timeline

## Signature Development

Signature Derivation/Validation

Algorithm Development/Validation

Clinical Testing Development

## Assay Development

LDT Protocol

CLIA/CAP Approval

## Approval & Reimbursement

Clinical Trials for alternative NET indications

Guidelines Approval

Medicare/ NGS Reimbursement

NETest

2027\*

NETest PPQ

2024

2027

PROSTest

Q4 2024

Partner

LuPSMA Predictor

Q1 2025

2025

Myeloma

COLONTest

Melanoma

LUNGTest

BREASTest

Pancreas

Wellness/RO test

2022-2024

Conceptual